

**Specification**

Sterile enrichment supplement to improve the growth and the aerotolerance of *Campylobacter* spp.

**Presentation**

	Packaging Details	Shelf Life	Storage
10 Freeze dried vials			
Vial	23x60 mm glass vials, tag labelled, White plastic cap -	49 months	2-25 °C
with: 3 ± 0.1 g	10 vials per box.		

**Composition**

Composition (g/vial)

Sodium pyruvate.....	0.125
Sodium bisulphite.....	0.125
Ferrous sulphate.....	0.125

**NOTE :** Each vial is sufficient to supplement  
500 ml of Agar Base for *Campylobacter* sps.

Reconstitute the original freeze-dried vial  
by adding  
Sterile Distilled Water..... 6 ml

**Description /Technique**Description:

The use of this supplement, added to Columbia Agar, Blood Agar Base No.2 or Campylobacter Agar Base, with 5-7% lysed defibrinated horse or sheep blood, allows a faster and easy way to determine *Campylobacter* spp. providing a better tolerance to the conditions of incubation.

The following addition of antibiotics permits the suppression of companion flora.

Technique:

Collect, dilute and prepare samples and volumes as required according to specifications, directives, official standard regulations and/or expected results.

Reconstitute the vial with the sterile diluent in aseptic conditions and add it to 500 ml of any melted Agar base cooled to 50°C, previously supplemented also with 5-7% lysed defibrinated horse or sheep blood.  
Do not overheat once supplemented.

Pour the complete medium into Petri dishes and, once solidified on a flat surface, spread the plates either by streaking or by spiral method.

Incubate the plates in microaerophilic conditions at 35 ± 2°C for 24-48h.

*Campylobacter* spp. best grown is at 42°C (except *Campylobacter fetus subsp. fetus*).

(Incubation times longer than those mentioned above or different incubation temperatures may be required depending on the sample or the specifications).

After incubation, count all the colonies that have appeared onto the surface of the agar.

Presumptive isolation of *Campylobacter* spp. must be confirmed by further microbiological and biochemical tests.

## Quality control

### Physical/Chemical control

Color : Orange      pH: at 25°C

### Microbiological control

Reconstitute 1 vial as indicated in COMPOSITION; shake and dissolve completely

Add 1 vial to 500 ml of medium base. DO NOT HEAT once supplemented.

Microaerophilia. Incubation at 35 ± 2°C or 42 ± 2°C during 24-48 horas

Analytical methodology according to ISO 11133:2014/A1:2018; A2:2020.

Distribute the complete medium, cooled to 50 °C, into 90 mm plates

Incubate according instructions for complete medium indicated in COMPOSITION.

Microaerophilic incubation at 35 ± 2 °C or 42± °C for 24-48 h

### **Microorganism**

*Campylobacter jejuni* ATCC® 29428, WDCM 00156

*Camp. coli-jejuni* ATCC® 33291, WDCM 00005

### **Growth**

Good

Good

### Sterility Control

Add 5 ml of the sample to:

100 ml TSB and 100 ml Thioglycollate.

Incubation 48 h at 30-35 °C and 48 h at 20-25 °C: NO GROWTH.

## Bibliography

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- CORRY, J.E.L., H.I. ATABAY, S.J. FORSYTHE & L.P. MANSFIELD (2003) Culture Media for the Isolation of *Campylobacters*, *Helicobacters* and *Arcobacters*, en Corry et al. (Eds) Handbook of Culture Media for Food Microbiology Chap 18 pgs 271-316. Elsevier Science B.V. Amsterdam.
- DIN 38411-6 (1991) Deutsche Einheitsverfahren zur Wasser-, Abwasser- und Schlammuntersuchung; Mikrobiologische Verfahren (Gruppe K); Nachweis von *Escherichia coli* und coliformen Keimen (K6)
- ISO 11133:2014/ Adm 1:2018. Microbiology of food, animal feed and water. Preparation, production, storage and performance testing of culture media.