

## Specification

Selective and differential medium used in the detection, isolation and enumeration of *Salmonella* and coliforms in clinical specimens according to the Pharmacopoeial Harmonized Methodology and in foodstuffs specimens according to ISO standard 21150.

## Presentation

30 Membrane filtration plates  
55 mm Plates for filtration purposes  
with: 9 ± 1 ml

### Packaging Details

1 box containing: 6 plastic bags with 5 plates of 55 mm/ bag.

### Shelf Life

6 months

### Storage

2-25 °C

## Composition

Composition (g/l):
Pancreatic digest of gelatin..... 17.000
Peptone of meat..... 1.500
Peptone of casein..... 1.500
Lactose monohydrate..... 10.000
Bile salts..... 1.500
Sodium chloride..... 5.000
Neutral red..... 0.030
Crystal violet..... 0.001
Agar..... 13,5

## Description /Technique

Collect, dilute and prepare samples and volumes to be filtered as required according to specifications, directives, official standard regulations and/or expected results.

Filter the sample through a 0.45 µm pore membrane and apply it onto the surface of the agar.

Incubate the plates right side up at 35+/-2°C for 24-48h.

After incubation, enumerate all the colonies that have appeared onto the surface of the filter.

Coliforms will develop reddish lactose-fermenting colonies. As violet crystal and ox bile inhibit gram-positive microorganisms, the growth of reddish colonies on plates incubated at 37°C will suggest total coliform presence in the sample; the growth of reddish colonies on plates incubated at 44°C will indicate faecal coliforms presence.

Calculate total microbial count per ml of sample by multiplying the average number of colonies per membrane by the inverse dilution factor. Report results as Colony Forming Unit (CFU's) per ml along with incubation time and temperature.

Each laboratory or industry must establish their own monitoring programme and evaluate its results according to their specifications.

Revision date: 10/10/24

## Quality control

### Physical/Chemical control

Color : Pink                            pH:  $7.1 \pm 0.2$  at  $25^\circ\text{C}$

### Microbiological control

Growth Promotion Test 50-100 CFU according to harmonized Pharmacopoeia monographs (EP) and test methods & ISO 11133:2014/A1:2018  
Membrane Filtration /Practical range  $100 \pm 20$  CFU. min. 50 CFU (productivity)./ $<100$  CFU (especificidad-PhEur) and  $\geq 10^3$  CFU (specificity-ISO)/ $10^4$ - $10^6$  CFU (selectivity)  
Analytical methodology according to ISO 11133:2014/A1:2018; A2:2020.

Aerobiosis. Incubation at  $35 \pm 2$  °C, reading at 24-48 hours.

*S. sonnei* incubation at  $37^\circ\text{C}$  for 20-24h

### Microorganism

*Enterococcus faecalis* ATCC® 19433, WDCM 00009

*Staphylococcus aureus* ATCC® 6538, WDCM 00032

*Escherichia coli* ATCC® 8739, WDCM 00012

*Escherichia coli* ATCC® 25922, WDCM 00013

*Salmonella typhimurium* ATCC® 14028, WDCM 00031

*Ps. aeruginosa* ATCC® 9027, WDCM 00026

*Shigella sonnei* ATCC® 9290

### Growth

Inhibited

Inhibited

Good ( $\geq 50\%$ ) - Red purple colonies - Biliar precipitate

Good ( $\geq 50\%$ ) - Red purple colonies - Biliar precipitate

Good ( $\geq 50\%$ ) - colourless colonies w/o precipitate

Colourless colonies without biliar precipitate

Good -colourless colonies w/o precipitate

### Sterility Control

Incubation 48 h at  $30$ - $35$  °C and 48 h at  $20$ - $25$  °C: NO GROWTH.

Check at 7 days after incubation in same conditions.

## Bibliography

- ATLAS, R.M., L.C. PARKS (1993) Handbook of Microbiological Media. CRC Press, Inc. London.
- CLESKERI, L.S., A.E. GREENBERG & A.D. EATON (1998) Standard Methods for the Examination of Water and Wastewater. 20th ed. APHA-AWWA-WEF. Washington. DC. USA.
- DOWNES, F.P. & K. ITO (2001) Compendium of Methods for the Microbiological Examination of Foods. 4th ed. APHA. Washington.
- EUROPEAN PHARMACOPOEIA 8.0 (2014) 8th ed. § 2.6.13. Microbiological examination of non-sterile products: Test for specified microorganisms. Harmonised Method. EDQM. Council of Europe. Strasbourg.
- HITCHINS, A.D., P. FENG, W.D. WATKINS, S.R. RIPEY & C.A. CHANDLER (1998) *E. coli* and coliform bacteria. Bacteriological Analytical Manual. 8th ed. AOAC International. Gaithersburg. MD. USA.
- HORWITZ, W. (2000) Official Methods of Analysis. AOAC Intl. Gaithersburg. MD. USA.
- ISO 11133:2014/ Adm 1:2018. Microbiology of food, animal feed and water. Preparation, production, storage and performance testing of culture media.
- ISO Standard 21567: 2004. Microbiology of Foods and animal feeding stuffs - Horizontal method for the detection of *Shigella* spp.
- ISO 4973:2023. Quality control of culture media and diluents used in cosmetics standards
- ISO 21150: 2015. Standard. Cosmetics - Detection of *E. coli*.
- MacCONKEY, A.T. (1905) Lactose-fermenting Bacteria in faeces. J. Hyg 5:333.
- MURRAY, P.R., E.J. BARON, M.A. PFALLER, F.C. TENOVER, & R.H. YOLKEN (Eds) (1995) Manual of Clinical Microbiology. 6th ed. A.S.M. Washington. DC. USA.
- RAPPAPORT, F. & E. HENING (1952) Media for the isolation and differentiation of pathogenic *E. coli* (serotypes O111 and O55) J. Clin. Pathology 5:361-362.
- USP 33 - NF 28 (2011) <62> Microbiological examination of non-sterile products: Test for specified microorganisms. Harmonised Method. USP Corp. Inc. Rockville. MD. USA.
- VARNAM, A.H. & M.G. EVANS (1991) Foodborne pathogens. Manson Publishing Ltd. London. UK.
- WHO (1963) International Standards for Drinking Waters. 7th ed. Churchill Ltd. London.