

Specification

Selective medium for the isolation of pathogenic staphylococci, according to the Pharmacopoeial Harmonized Methodology and Clinical samples.

Presentation

	Packaging Details	Shelf Life	Storage
30 Membrane filtration plates 55 mm Plates for filtration purposes with: 9 ± 1 ml	1 box containing: 6 plastic bags with 5 plates of 55 mm/ bag.	6 months	2-25 °C

Composition

Composition (g/l):

Meat Peptone.....	5.000
Beef extract.....	1.000
Peptone from casein	5.000
Sodium chloride.....	75.000
D(-)Mannitol.....	10.000
Phenol red.....	0.025
Agar.....	15.000

Description /Technique

Description:

Mannitol Salt Agar is a classical medium for the detection and enumeration of staphylococci. It was described by Chapman and has been adopted by many official organisations. Several modifications of it have been developed, all formulations resulting in media with similar efficiency.

This medium takes advantage of the high saline tolerance of staphylococci, and uses sodium chloride as a selective agent. Only staphylococci and halophilic enterobacteria are able to grow freely at the concentration of salt employed in this medium, while other bacteria are inhibited. It also exploits the correlation between the pathogenicity of staphylococci and their ability ferment mannitol. Mannitol fermentation results in an accumulation of acid products, indicated by the phenol red indicator turning yellow. A yellow halo surrounds the presumptive pathogenic colonies, while the rest of the medium remains red/orange in colour.

Technique:

Collect, dilute and prepare samples and volumes as required according to specifications, directives, official standard regulations and/or expected results.

Filter the sample through a 0.45 µm pore membrane and apply it onto the surface of the agar.

Incubate the plates aerobically at 30-35 °C for 24-48-72 h

(Incubation times longer than those mentioned above or different incubation temperatures may be required depending on the sample, on the specifications,...)

After incubation, enumerate all the colonies that have developed onto the surface of the membrane, considering both yellowish colonies and red or colourless colonies.

Calculate total microbial count per ml of sample by multiplying the average number of colonies per plate by the inverse dilution factor .

Report results as Colony Forming Unit (CFU's) per ml along with incubation time and temperature.

Presumptive isolation of *S. aureus* must be confirmed by further microbiological and chemical tests.

Note: According to the methodology chosen by the laboratory (Pharmacopoeia or other international standards), may be slight variations in incubation times and temperatures, as well as inhibition of *E. coli*, which can be variable depending on the inoculated bacterial population. This medium can normally reduce the bacterial load by up to 3 decimal logarithms.

Quality control

Physical/Chemical control

Color : Strongly pink pH: 7.4 ± 0.2 at 25°C

Microbiological control

Membrane Filtration. Inoculate with 10-100 CFU according to harmonized Pharmacopoeiae or with 100-1000 CFU for selectivity.

Analytical methodology according to ISO 11133:2014/A1:2018; A2:2020.

Aerobiosis. Incubation at 30-35°C. Reading at 18-72h

Microorganism

Staphylococcus aureus ATCC® 6538, WDCM 00032

Stph. aureus ATCC® 25923, WDCM 00034

Stph. epidermidis ATCC® 12228, WDCM 00036

Escherichia coli ATCC® 8739, WDCM 00012

Growth

Good (≥50 %)

Good (≥50 %)

Poor to good

Inhibited

Sterility Control

Incubation 48 h at 30-35 °C and 48 h at 20-25 °C: NO GROWTH.

Check at 7 days after incubation in same conditions.

Bibliography

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